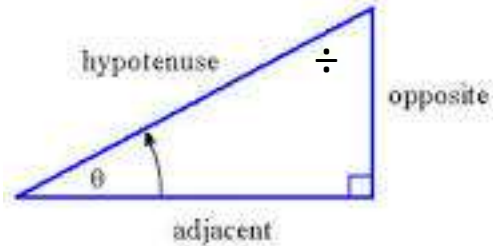
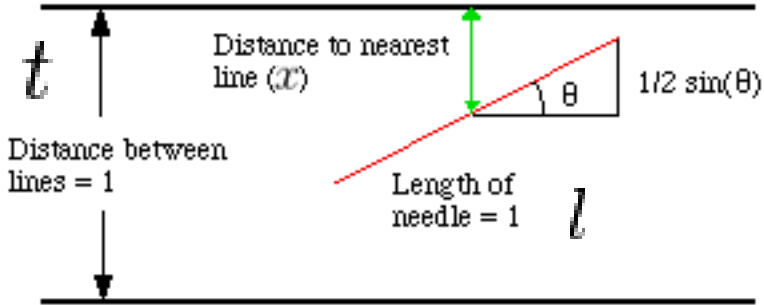


Molecular Dynamics
&
PELE

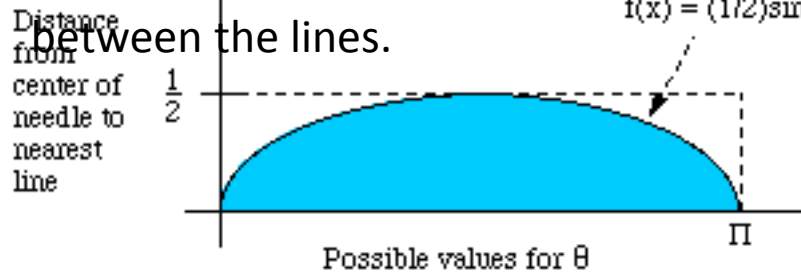
What is the Probability the Needle Crosses a Line?

- The 1st Monte Carlo Experiment

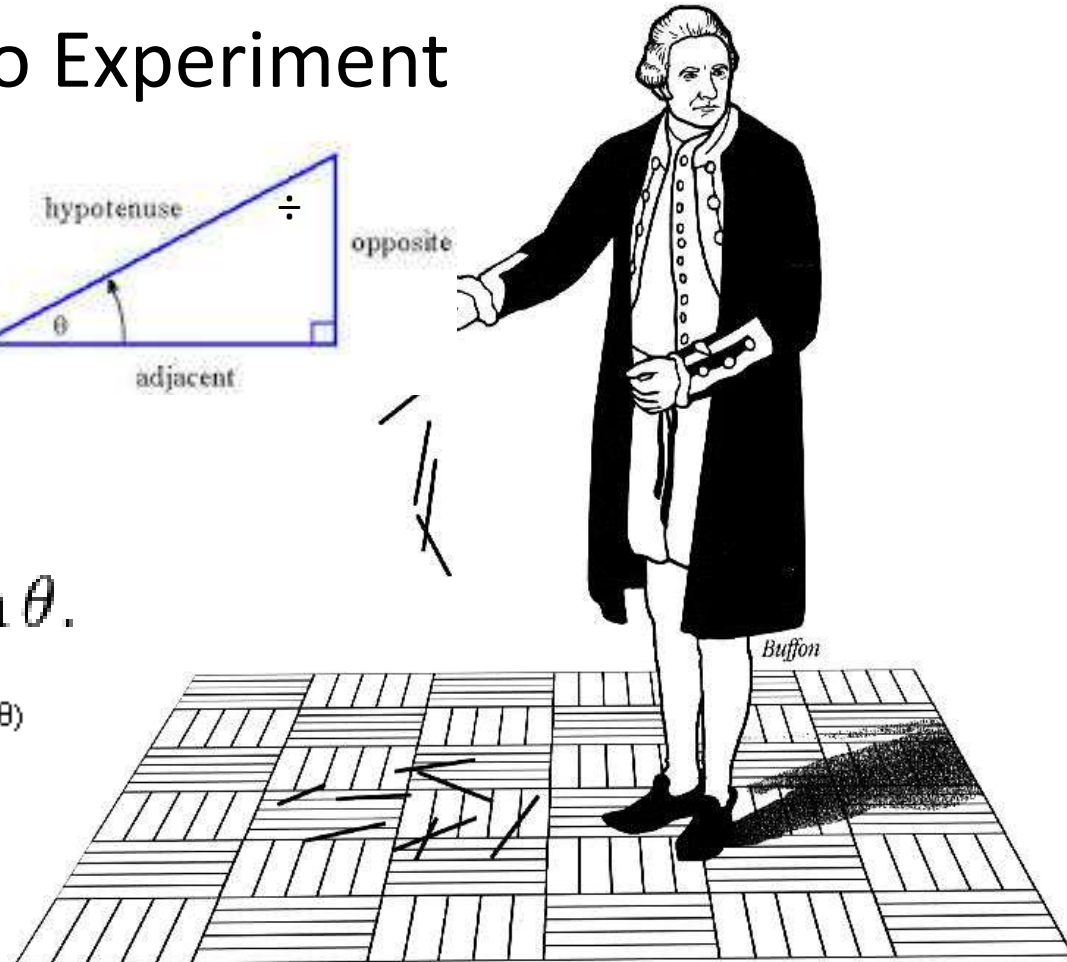


The distance from the center to the closest line can never be more than half the distance between the lines.

$$x \leq \frac{l}{2} \sin \theta.$$

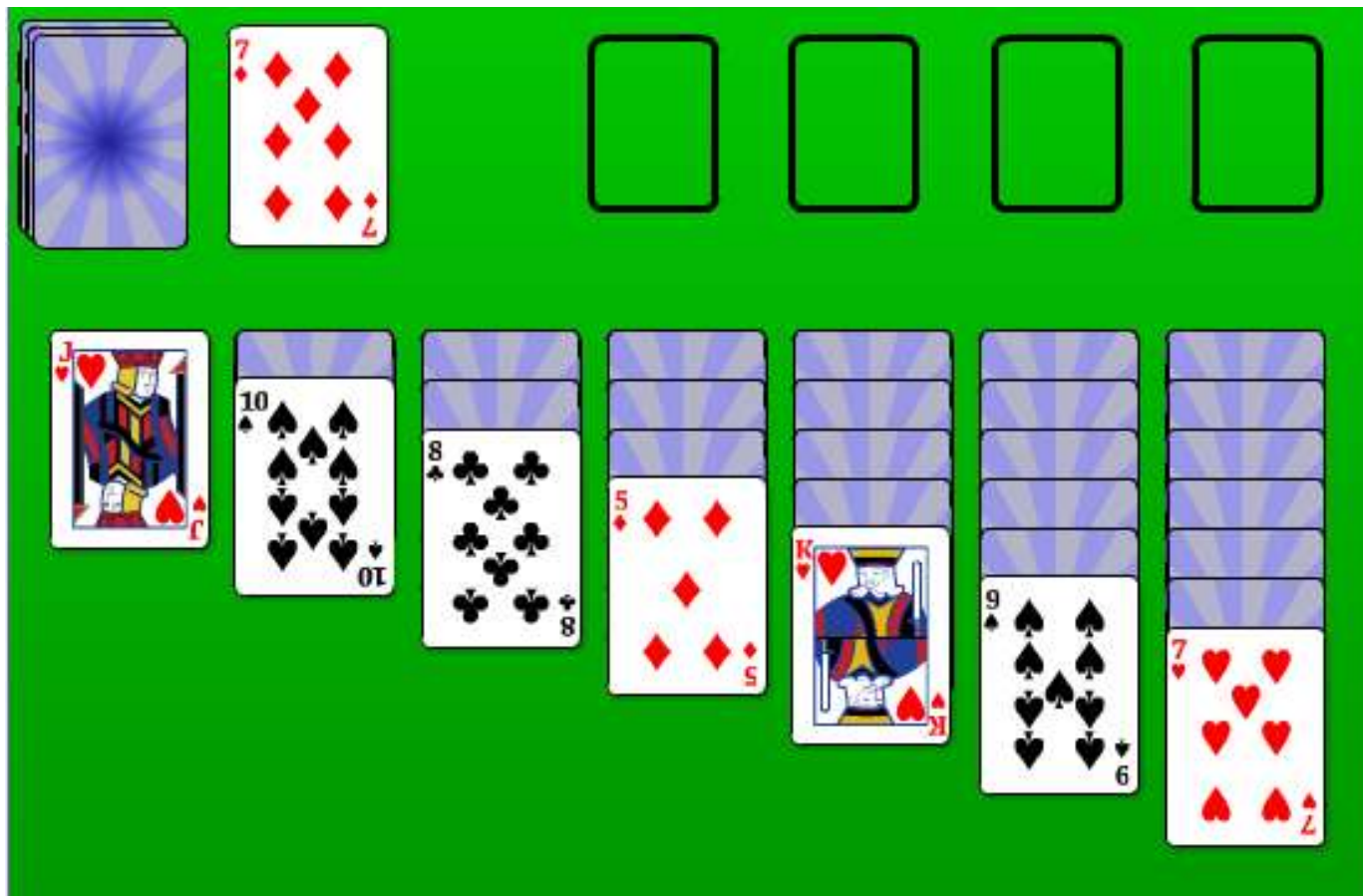


$$2(n)/th \approx \pi$$



Georges-Louis Leclerc, Comte de Buffon (1777)

Combinatorial Analysis & Theory of Probabilities



Metropolis Monte Carlo

1. Initial atom coordinates.
2. Random atom i moves by random displacement: ΔX_i , ΔY_i , and ΔZ_i
3. Calculate the change of potential energy ΔV corresponding to this displacement.
4. If $\Delta V < 0$ accept the new coordinates and go to step 2.
5. Otherwise, if $\Delta V \geq 0$, select a random number R in the range $[0,1]$ and:
 - A. If $e^{-\Delta V/kT} < R$ accept the new coordinates and go to step 2,
 - B. If $e^{-\Delta V/kT} \geq R$ keep the original coordinates and go to step 2.

Phase Transition for a Hard Sphere System

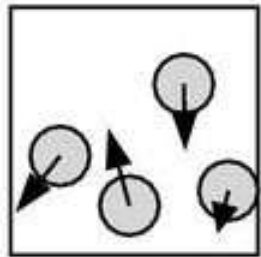
B. J. ALDER AND T. E. WAINWRIGHT

University of California Radiation Laboratory, Livermore, California

(Received August 12, 1957)

J. Chem. Phys. 27, 1208

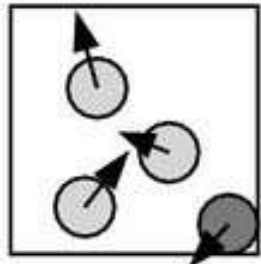
wall collision



$t = 0$



$t =$



$t = 4.04$



$t =$



03

.37

articles

Dynamics of folded proteins

J. Andrew McCammon, Bruce R. Gelin & Martin Karplus

Department of Chemistry, Harvard University, Cambridge, Massachusetts 02138

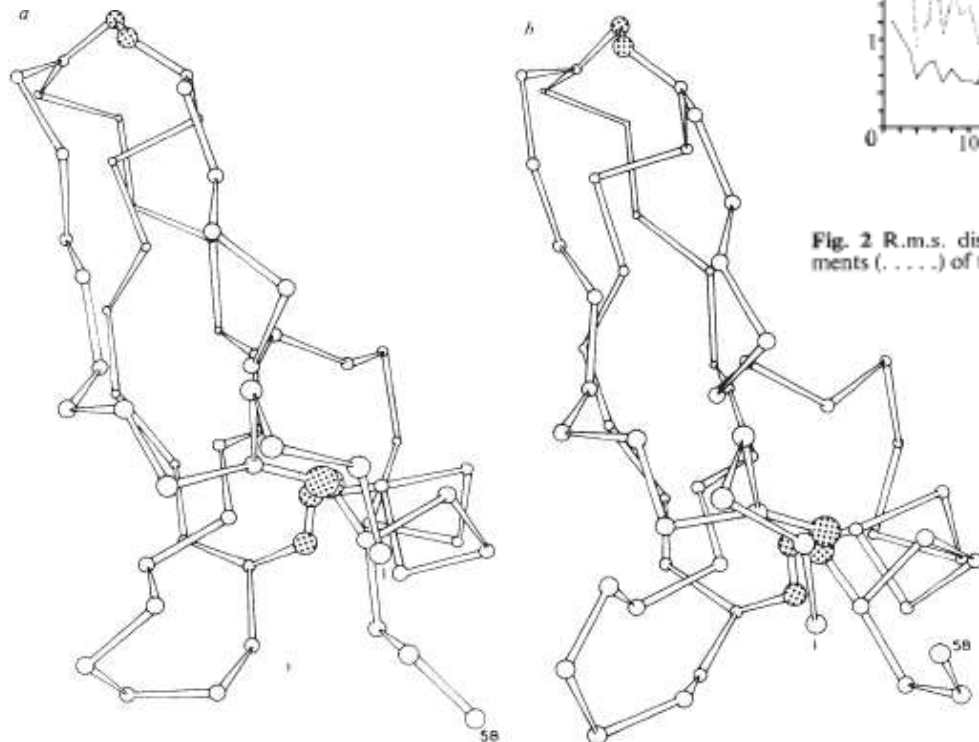


Fig. 1 The peptide backbone (α carbons) and disulfide bonds of BPTI. *a*, X-ray structure¹¹. *b*, Time evolved structure after 3.2 ps of dynamical simulation.

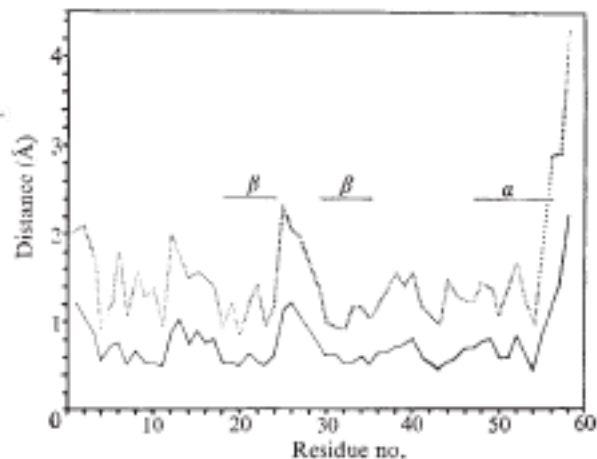


Fig. 2 R.M.S. displacements (—) and maximum displacements (....) of the α carbons, relative to the dynamical average structure.

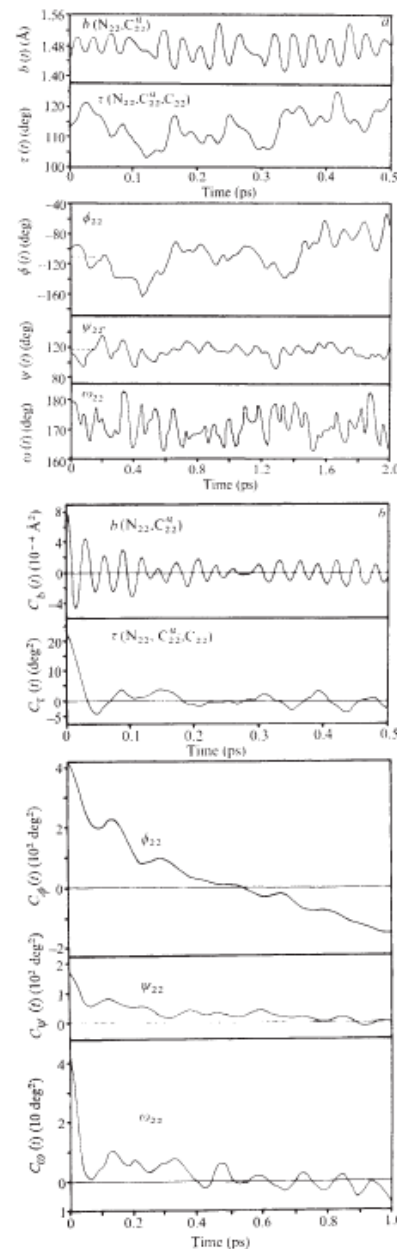
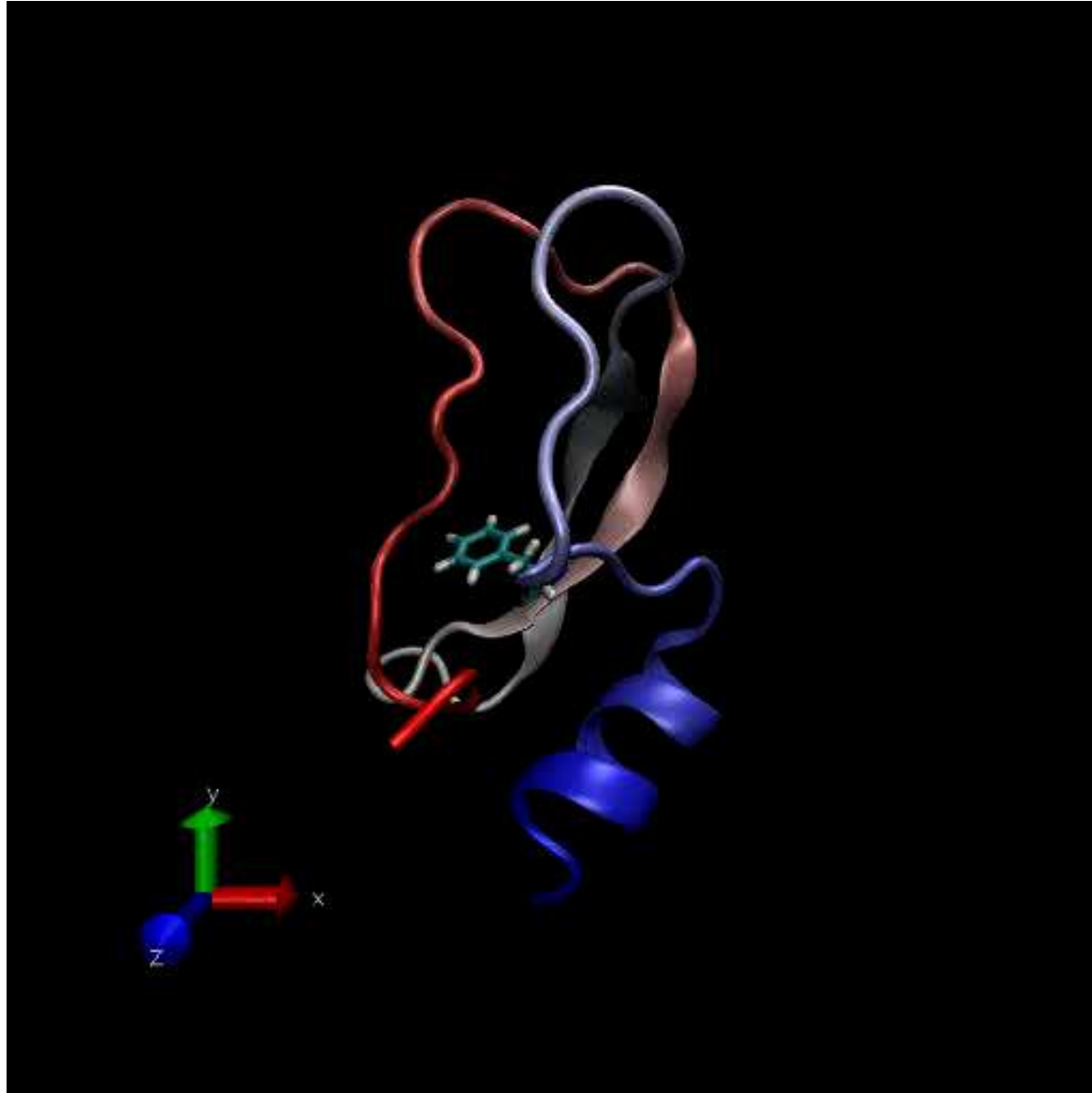


Fig. 3 *a*, Time development of selected Phe 22 internal coordinates; the initial time is 4.9 ps after the beginning of the dynamical simulation; average values are indicated by broken line segments. *b*, Time correlation functions for fluctuations of selected Phe 22 internal coordinates.

NMR of BPTI



Molecular dynamics simulations must pass through all possible states corresponding to the particular thermodynamic constraints

Molecular Dynamics

Statistical Mechanics

&

Classical Mechanics

Molecular Dynamics

- A computational method that calculates the time dependent behavior of a molecular system
- Provides detailed information on the fluctuations and conformational changes of ligands, proteins and nucleic acids

Definitions

- *Thermodynamic state* is usually defined by the temperature (**T**), the pressure (**P**), and the number of particles (**N**).
- The *mechanical or microscopic state* is defined by the atomic positions, (**r**), and momenta, (**p**); these can also be considered as coordinates in a multidimensional space called **phase space**.

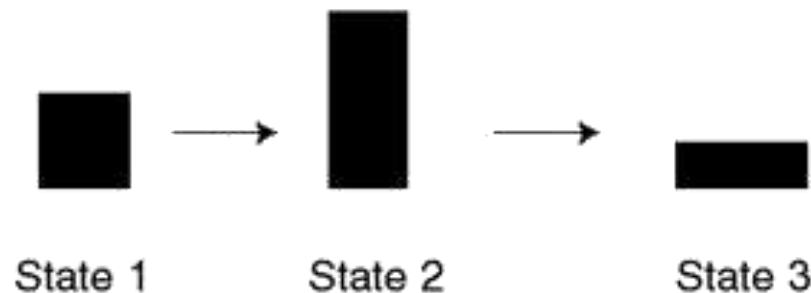
Molecular Dynamics

- An ensemble is a collection of all possible systems which have different microscopic states but have an identical macroscopic or thermodynamic state.

Thermodynamics describes the driving force for chemical processes

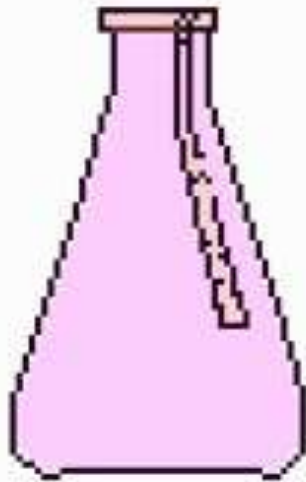


Kinetics describes the mechanism for the chemical process



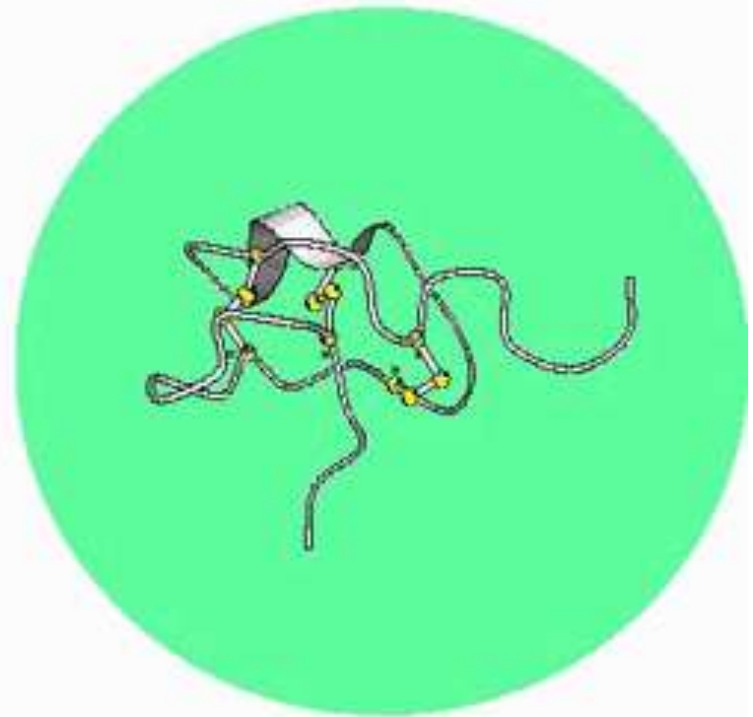
Molecular Dynamics

Experiment



Macroscopic

Molecular Simulation



Microscopic

Molecular Dynamics

- Statistical mechanics

Ensemble average = Time average

$$\langle A \rangle_{ensemble} = \iint dp^N dr^N A(p^N, r^N) \rho(p^N, r^N)$$

$\underbrace{\hspace{10em}}_{\text{Th}} \underbrace{\hspace{10em}}_{\text{pc}}$

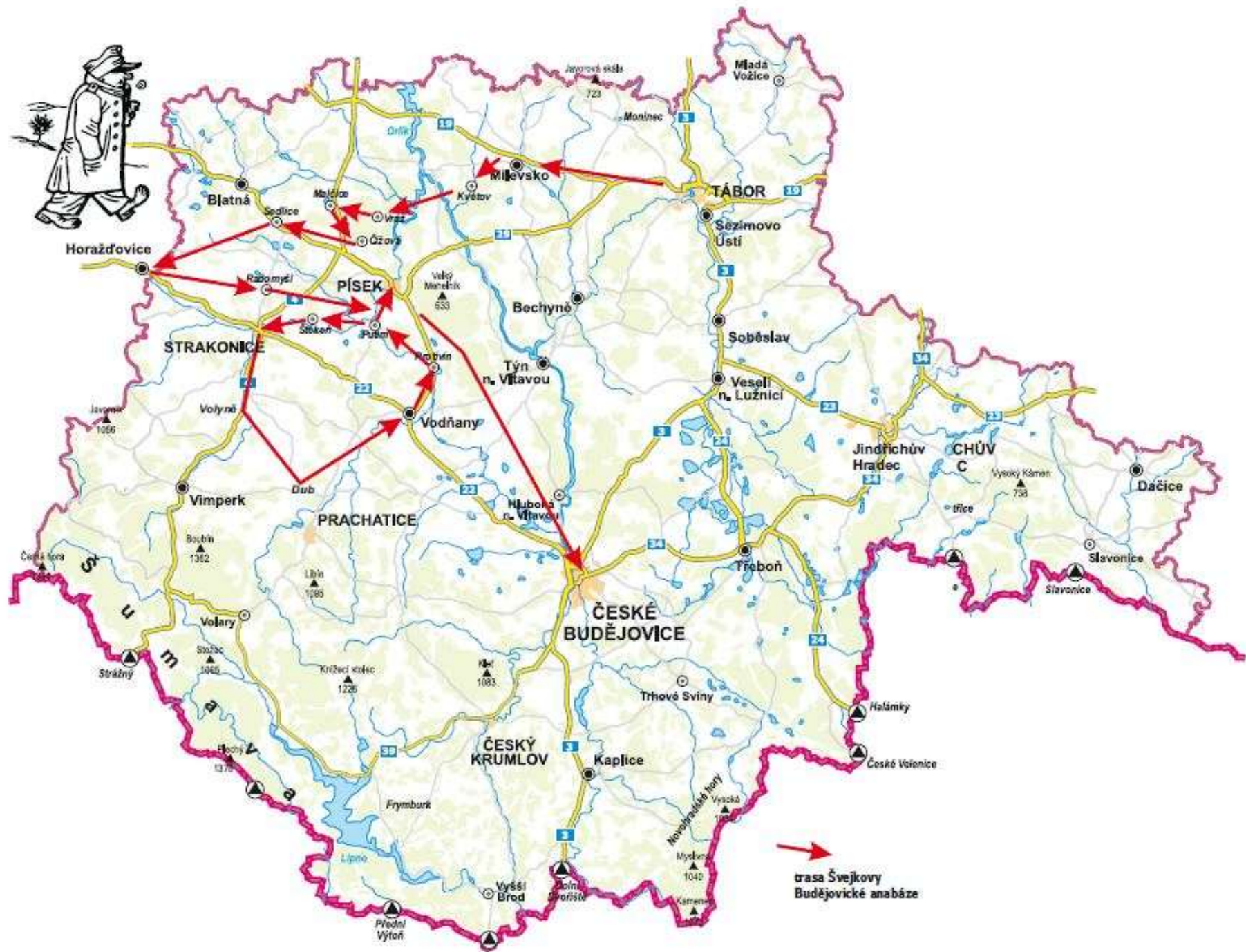
$$\rho(p^N, r^N) = \frac{1}{Q} \exp[-H(p^N, r^N) / k_B T]$$

momenta (p) and the positions ↓
 ↓ ↓
 partition function the thermodynamic state variables (temperature and volume)

$$\langle A \rangle_{time} = \lim_{\tau \rightarrow \infty} \frac{1}{\tau} \int_{t=0}^{\tau} A(p^N(t), r^N(t)) dt \approx \frac{1}{M} \sum_{i=1}^M A(p^N, r^N) = \frac{R}{N_A}$$

The average simulation time (t) is the number of time steps (M) in the simulation and the instantaneous value of A ($A(p^N, r^N)$)

The average simulation time (t) is the number of time steps (M) in the simulation and the instantaneous value of A ($A(p^N, r^N)$)

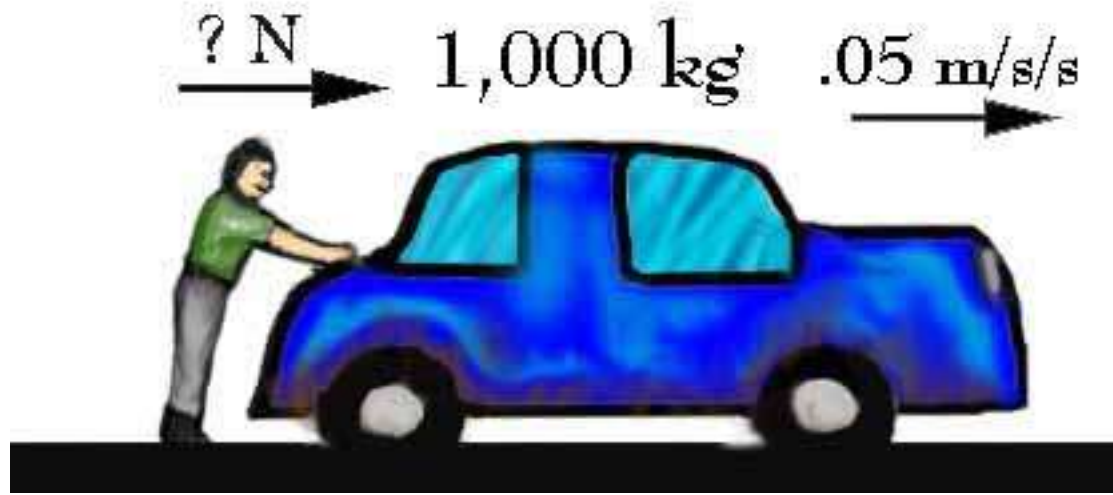


Molecular Dynamics

- **Ergodic hypothesis** = over a long period of time, the time spent by a system in some region of the phase space of microstates with the same energy is proportional to the volume of this region

Molecular Dynamics

- Classical mechanics
 - Newton's 2nd Law of Thermodynamics
 - $F=ma$ (the acceleration of an object is dependent on the net force acting upon the object and the mass of the object)



Molecular Dynamics

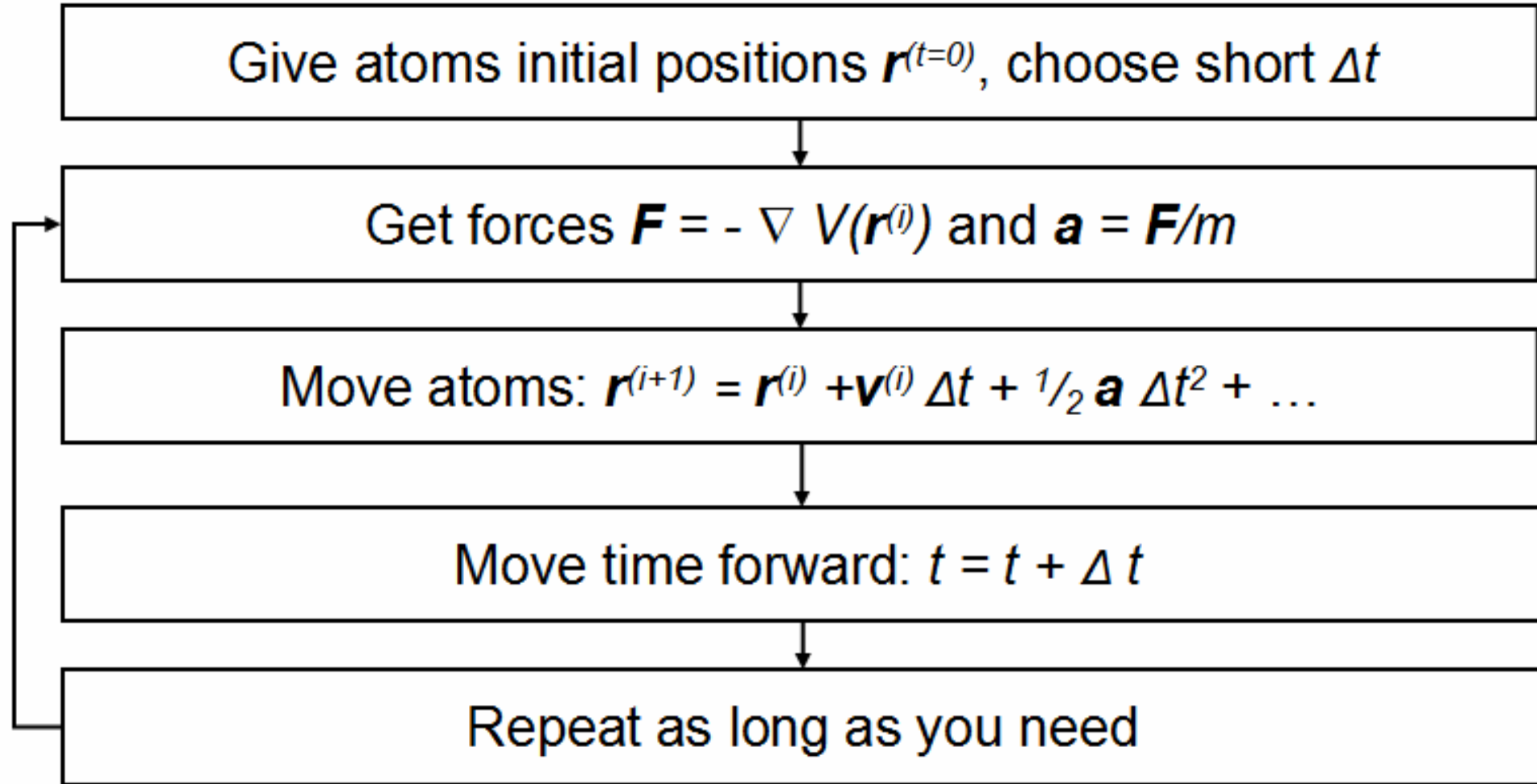
Give atoms initial positions $\mathbf{r}^{(t=0)}$, choose short Δt

Get forces $\mathbf{F} = -\nabla V(\mathbf{r}^{(i)})$ and $\mathbf{a} = \mathbf{F}/m$

Move atoms: $\mathbf{r}^{(i+1)} = \mathbf{r}^{(i)} + \mathbf{v}^{(i)} \Delta t + \frac{1}{2} \mathbf{a} \Delta t^2 + \dots$

Move time forward: $t = t + \Delta t$

Repeat as long as you need



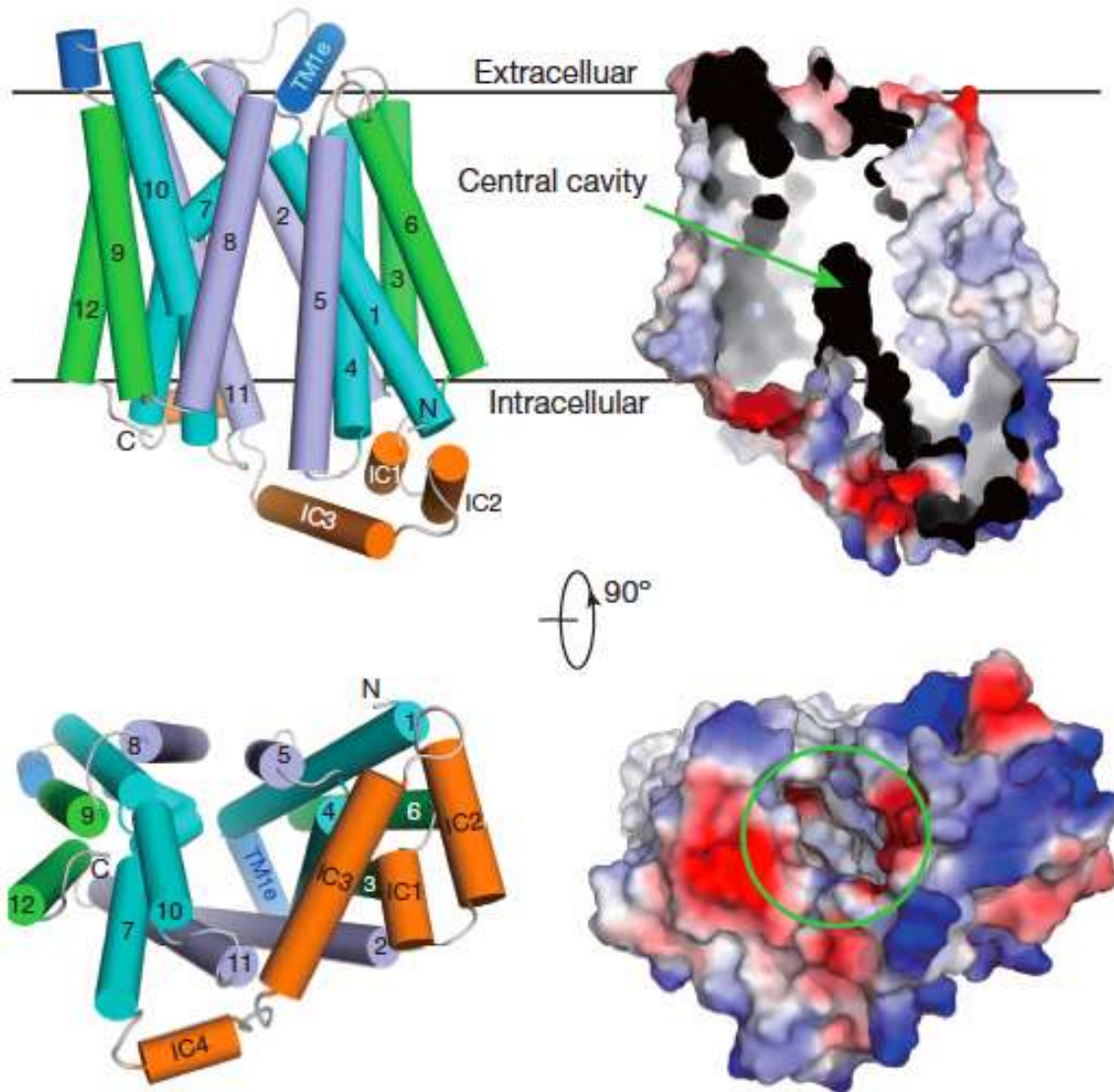
Application

Glucose Transporter System

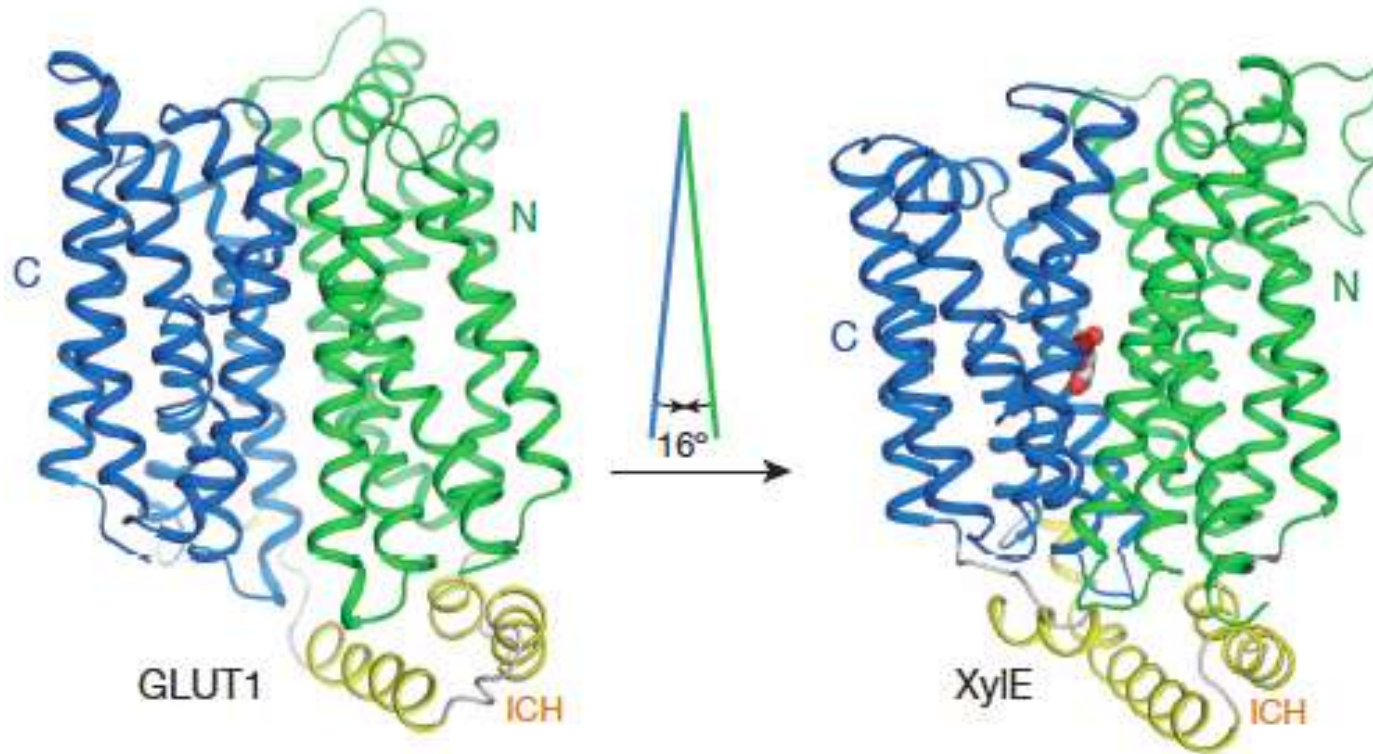
Crystal structure of the human glucose transporter GLUT1

2014 | VOL 510 | NATURE | 121

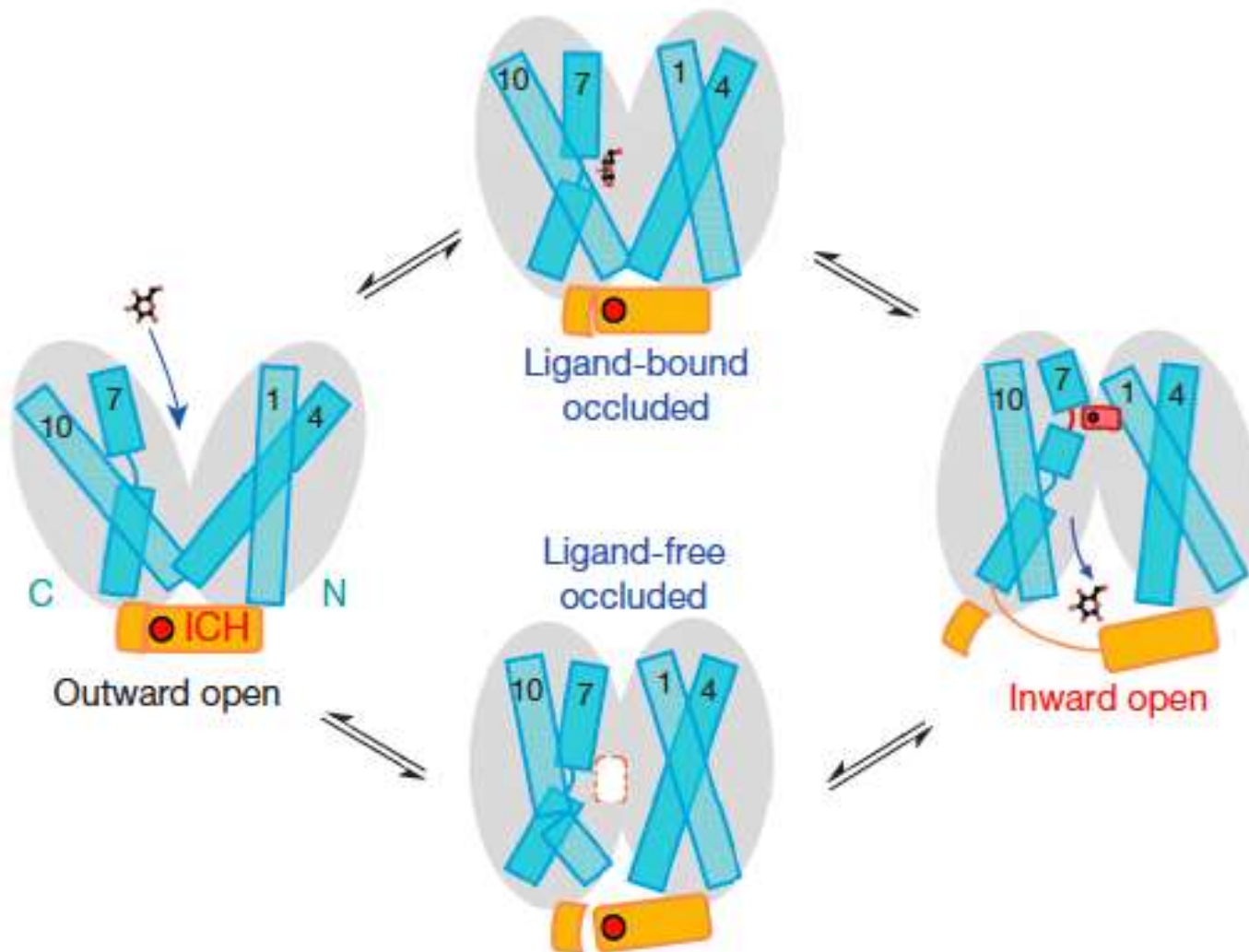
Dong Deng^{1,2,3*}, Chao Xu^{1,2,3*}, Pengcheng Sun^{1,2*}, Jianping Wu^{1,2,3*}, Chuangye Yan^{1,2}, Mingxu Hu^{1,2,3} & Nieng Yan^{1,2,3}



Structural Conformations



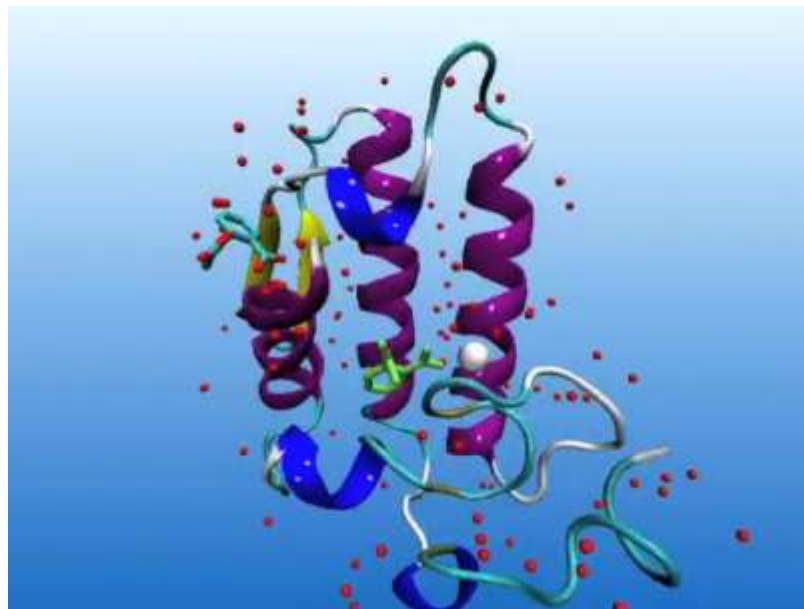
Structural Conformations



PELE

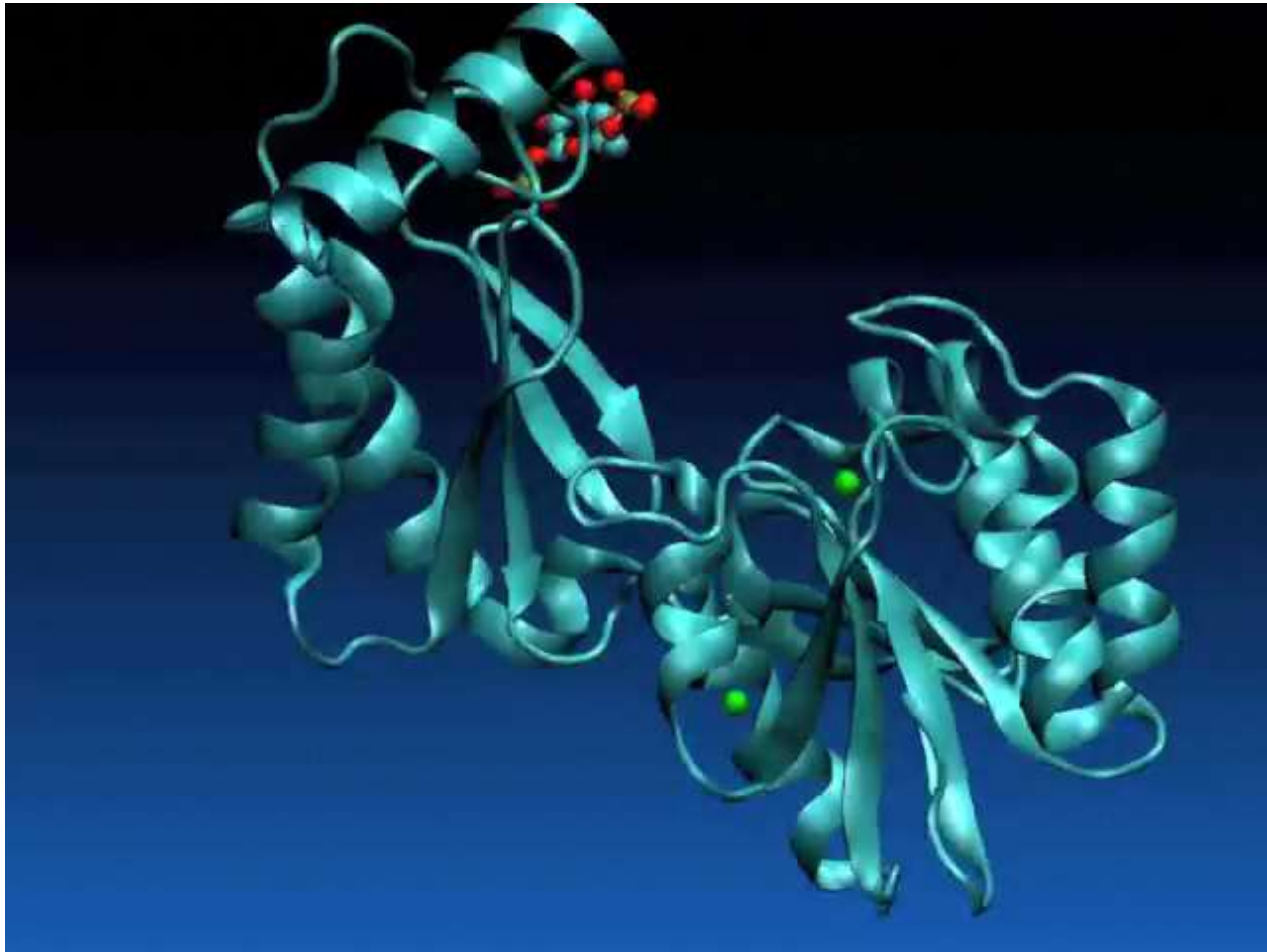
A Metropolis Monte-Carlo Software

PELE (PROTEIN ENERGY LANDSCAPE EXPLORATION)

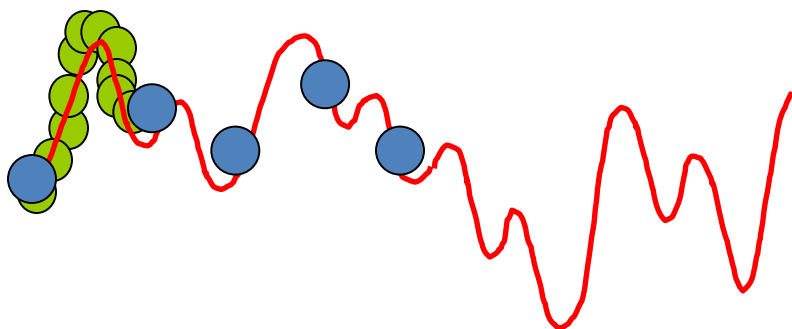


- Kenneth W. Borrelli, Andreas Vitalis, Raul Alcantara and Victor Guallar. Protein energy landscape exploration. A novel Monte Carlo technique. Implications on camphor ligand binding. *Journal of Chemical Theory and Computation*, **6**:1304-1311, (2005)
- Kenneth Borrelli, Benjamin Cossins, and Victor Guallar, Exploring hierarchical refinement techniques for induced fit docking with protein and ligand flexibility *J. Comp. Chem.*, **31**: 1224-35 (2010)
- Benjamin Coussins, Ali Hosseini and Victor Guallar, Exploration of Protein Conformational Change with PELE and Meta-Dynamics, *Journal of Chemical Theory and Computation*, **8**:959-965 (2012)
- Fatima Lucas and Victor Guallar, An Atomistic View on Human Hemoglobin Carbon Monoxide Migration Processes, *Biophysical Journal*, **102**:887-896 (2012)
- Hosseini, A., Espona-Fiedler, M., Soto-Cerrato, V., Quesada, R., Pérez-Tomás, R. & Guallar, V. Molecular Interactions of Prodiginines with the BH3 Domain of Anti-Apoptotic Bcl-2 Family Members. *PloS one* **8**, e57562 (2013)
- Takahashi, R., Gil, V.A. and Guallar, V., Monte Carlo Free Ligand Diffusion with Markov State Model analysis and Absolute Binding Free Energy Calculations. *Journal of Chemical Theory and Computation*, **10**:282-288. (2014)

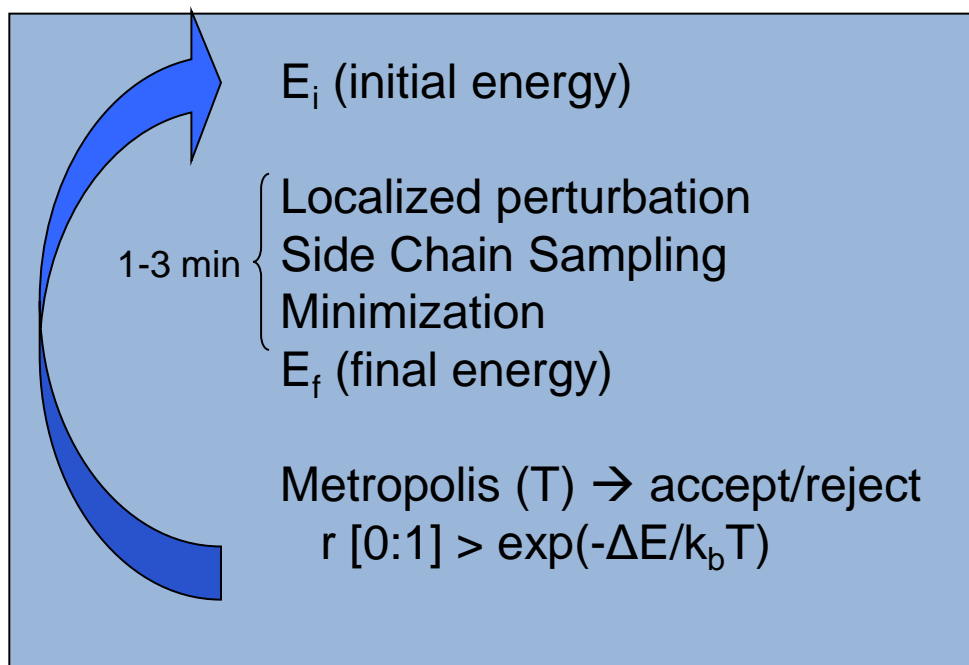
You collect what you input!



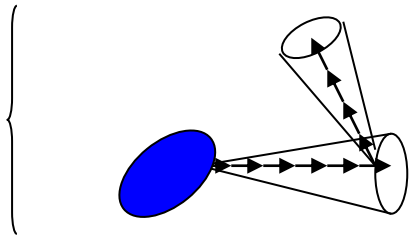
PELE: combining Monte Carlo with protein prediction algorithms



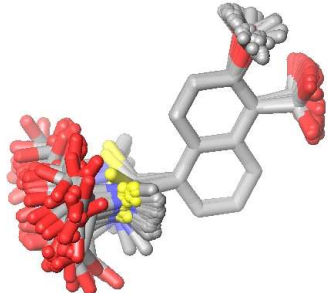
The algorithm (PELE):



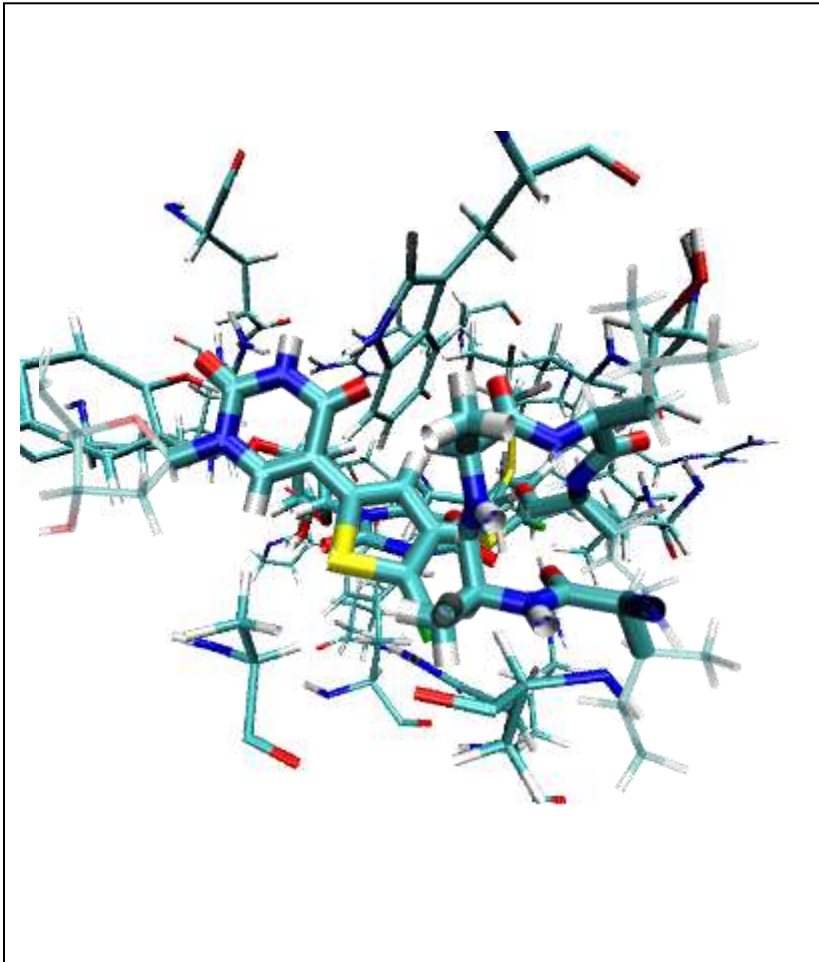
Steered the perturbation



Ligand motion



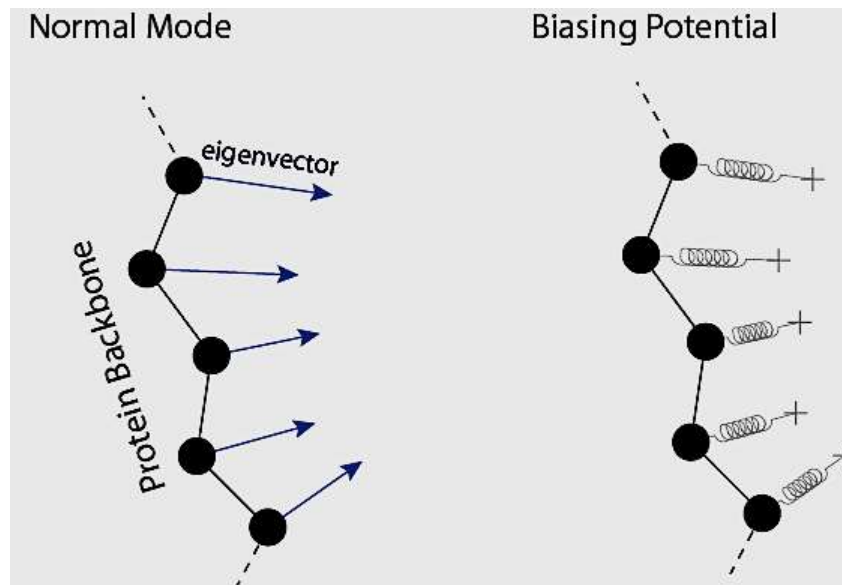
- Eliminate grid-points which always produce steric clashes (0.65% of VdW radius)
 - That create a steric clash between the ligand core and the protein sidechain
 - For which all the rotamers of a single residue or ligand region cause a steric clash with the backbone or ligand core
 - For which a single combination of rotamers cannot be found that does not produce a steric clash



Backbone Perturbation: Anisotropic Network Model

Follow a mode in an steered minimization

This ligand perturbation is coupled to backbone motion by means of a gaussian network of alpha carbon contacts



Side Chain Step (Prediction)

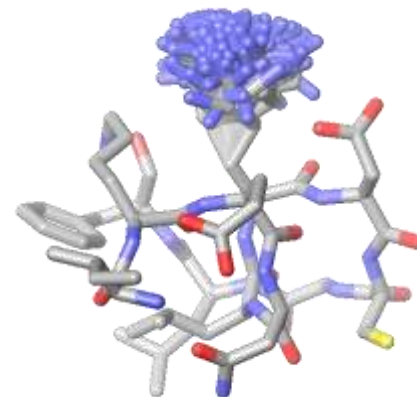
Rotamer Libraries (Barry Honig 2001) Define All Possible Residue Movements

Steric Filtering Removes Unfeasible Ones

Resulting Rotamers are Clustered

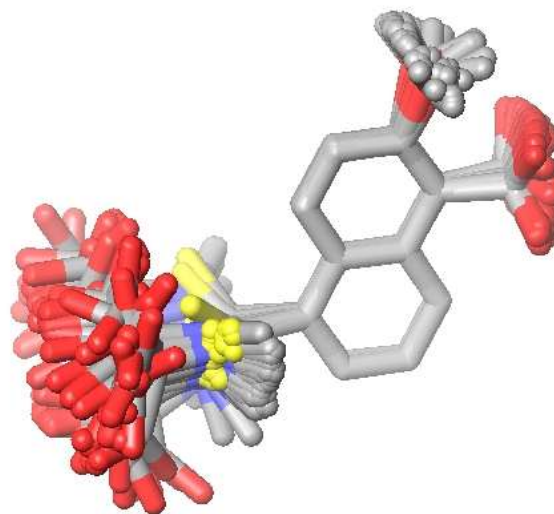
Clusters are Minimized

Lowest Energy Cluster is Selected



The Ligand also has rotamers associated!!

- Choosing a “core” region
 - The frozen region (no rotatable bonds within the region) that minimizes the number of bond lengths from this region to the farthest atom
- Populate the Library
 - Using Macromodel: eliminating all those that create a steric clash (two atoms closer than 75% of their combined VdW distance)



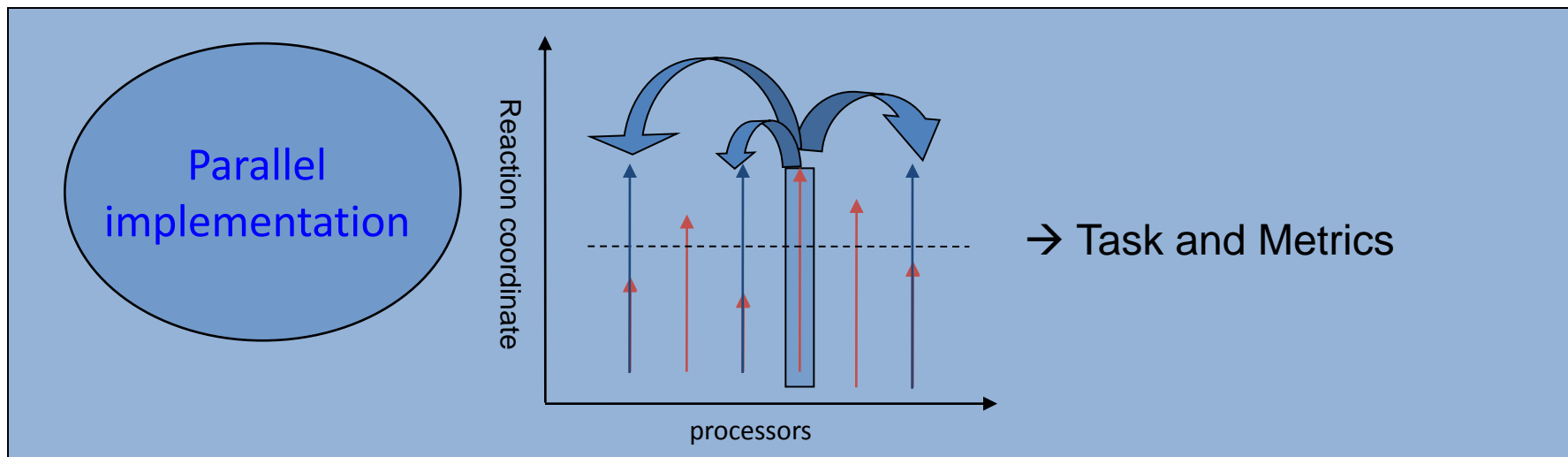
3 independent rotamers

Last step: minimization

Truncated Newton with long/short range force update and numerical born alpha derivatives

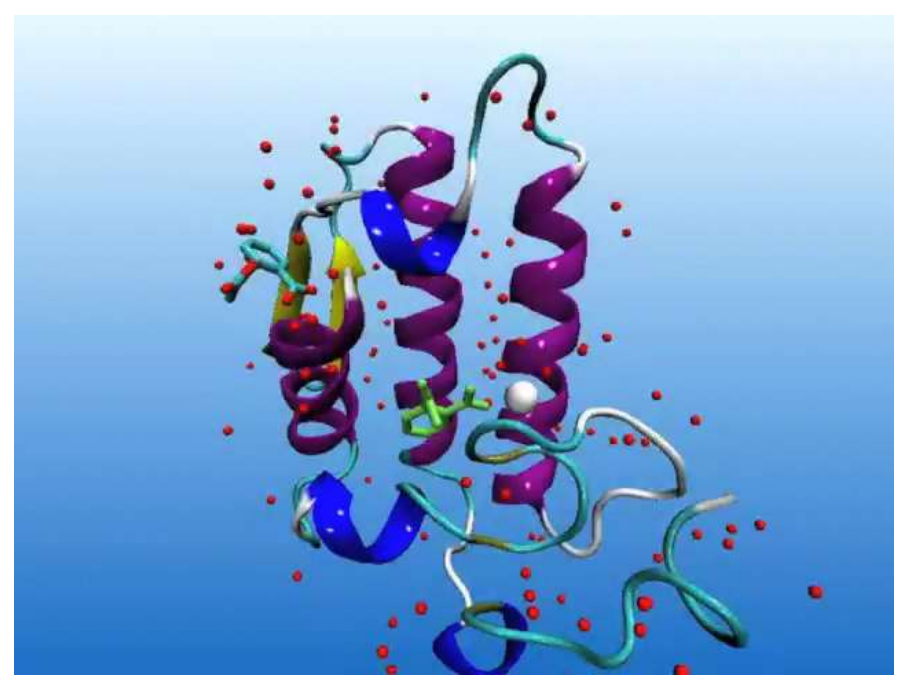
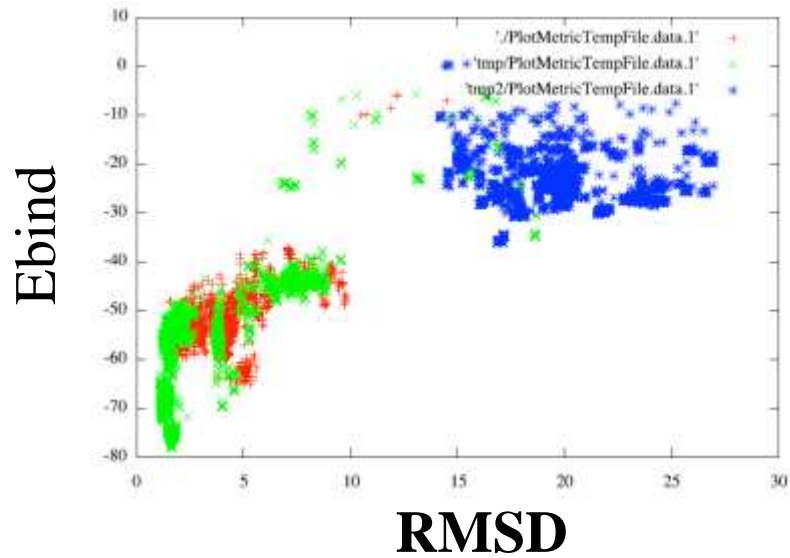
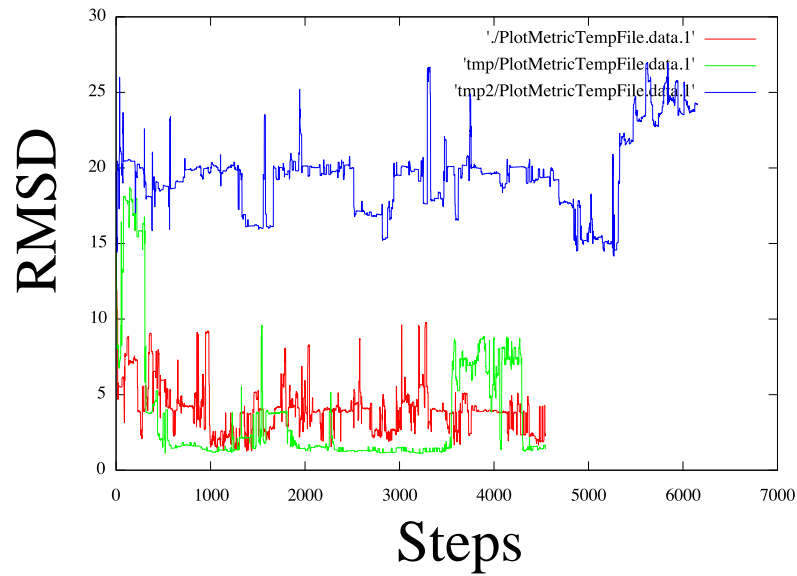
Alpha carbon can/should be constrained to the outcome of ANM

All rap up in MPI (parallel implementation: one node controlling and n-1 running)

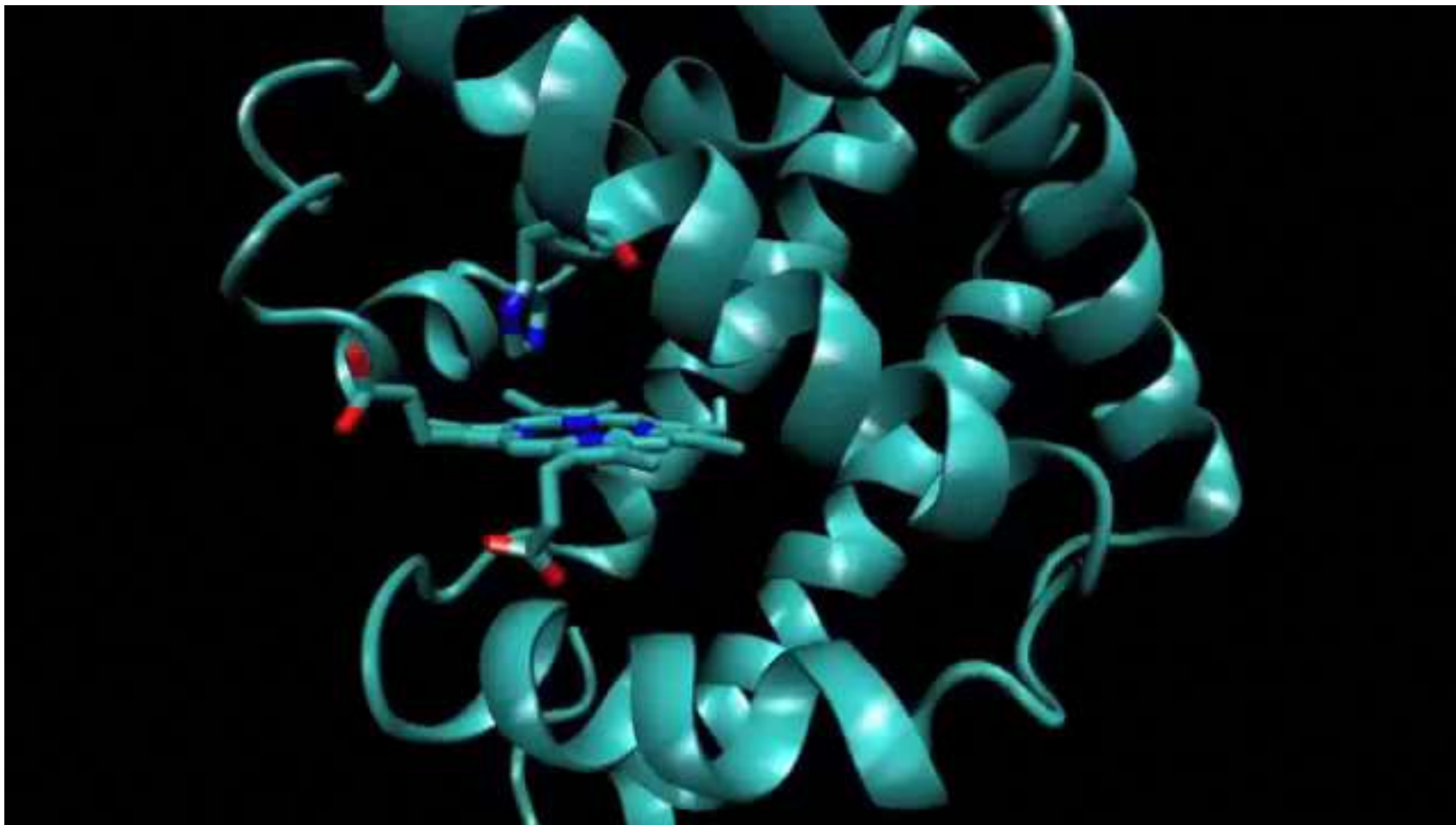


Tasks: lower energy, binding energy, move ligands away/to, move domains, change initial rmsd, etc.

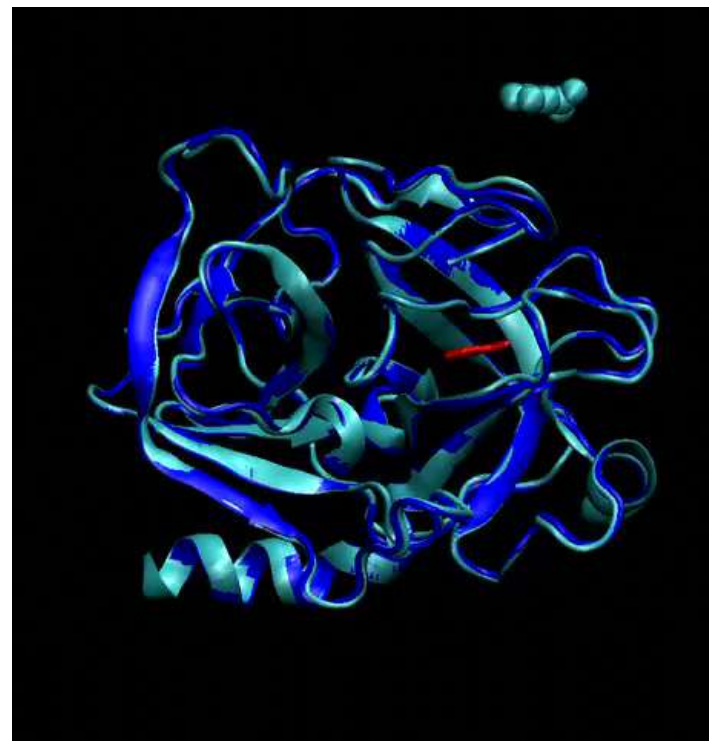
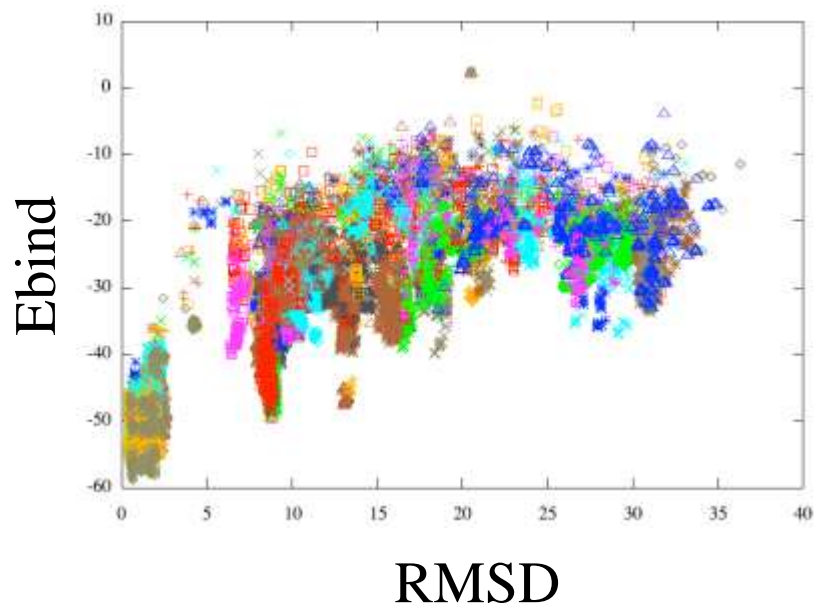
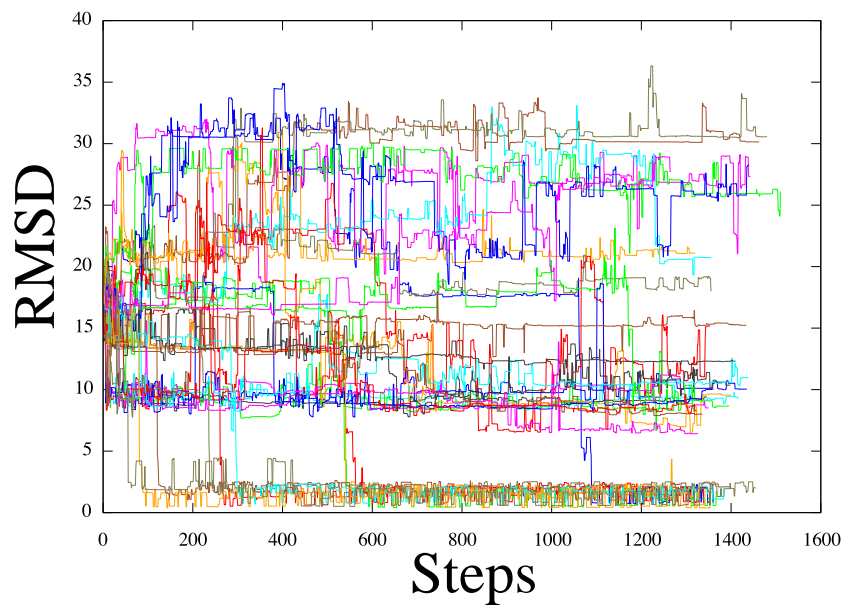
Free search with PELE



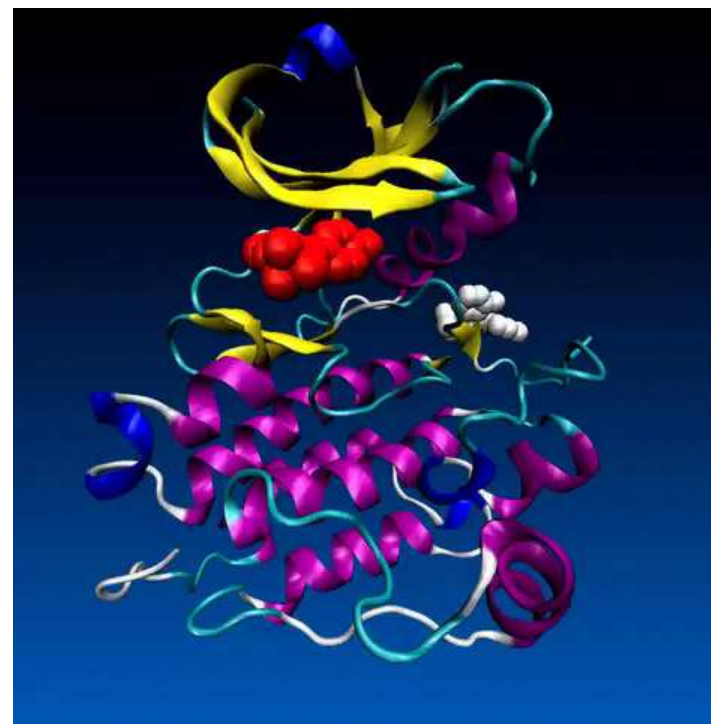
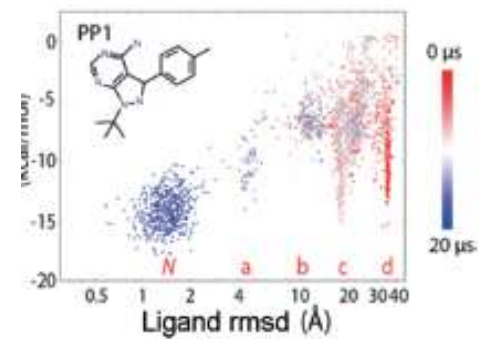
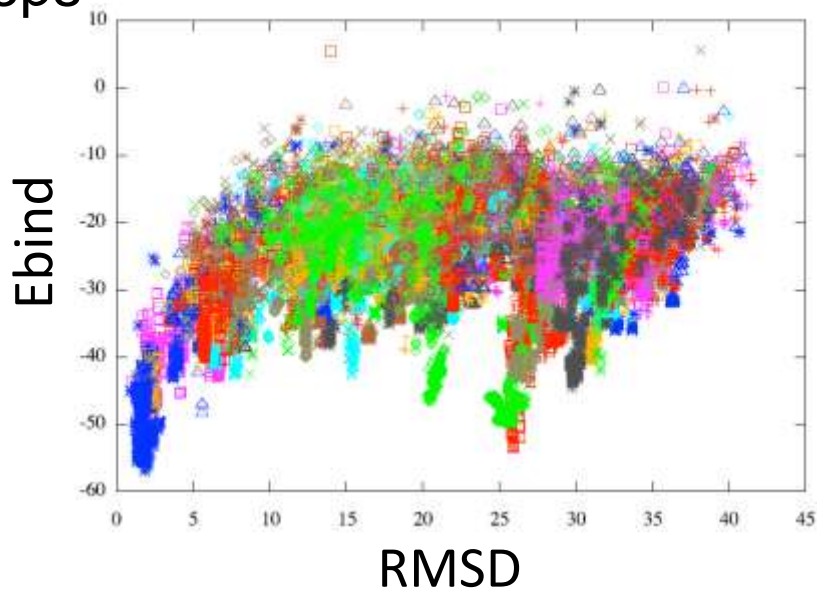
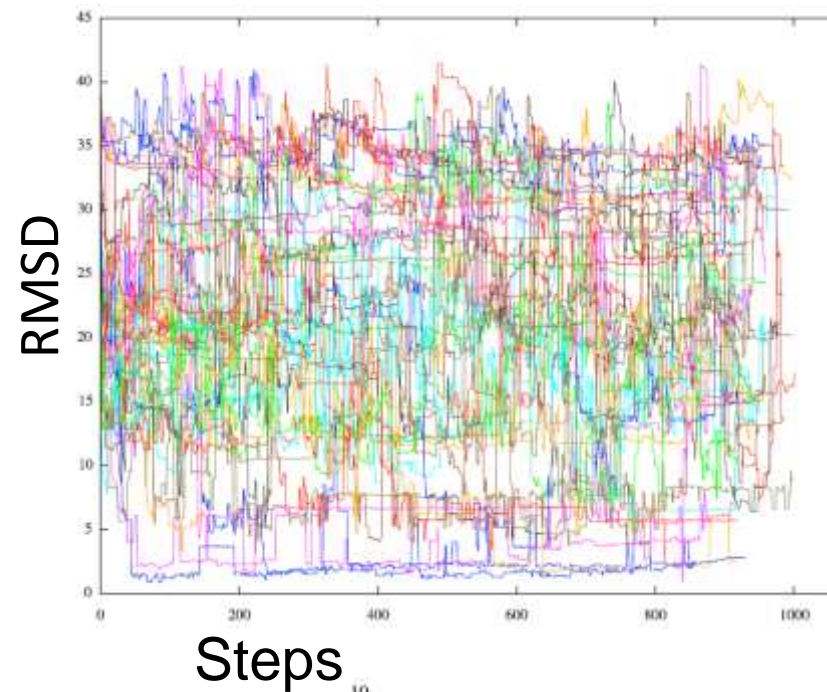
Want to see carbon monoxide getting inside Mb?



Full ligand random exploration: Benzamide binding to the bovine trypsin.



Can we do a difficult case like the Src Kinase?



The PELE Server

The screenshot shows a web browser window with the URL <https://pele.bsc.es/pele.wt/about>. The page header features the BSC logo (Barcelona Supercomputing Center) and the PELE logo. A navigation menu on the left includes links for About, News, Help, Register, and Login. The main content area is titled "About PELE" and contains a paragraph describing the method, a list of three steps (Localized perturbation, Side Chain Sampling, and Minimization), and a diagram illustrating the energy landscape exploration process. The diagram shows a blue arrow curving from an initial energy state E_i to a final energy state E_f , with a bracket indicating a duration of 1-2 minutes for the intermediate steps.

Barcelona Supercomputing Center
Centro Nacional de Supercomputación

PELE

0.9.6.1

- About
- News
- Help
- Register
- Login

About PELE

Using technological advances in protein structure prediction, we have developed **PELE** (an acronym for **P**rotein **E**nergy **L**andscape **E**xploration), a novel method to perform protein energy landscape explorations. **PELE** combines a Monte Carlo stochastic approach with protein structure prediction algorithms and is capable of accurately reproducing long time scale processes in only few hours of CPU.

Figure below shows the heuristic algorithm for the landscape exploration method which is based on three main steps: *an initial perturbation*, *a side chain sampling* and *a final minimization*.

The diagram illustrates the heuristic algorithm for landscape exploration. It shows a blue arrow curving from an initial energy state E_i (initial energy) to a final energy state E_f (final energy). The process is divided into three main steps: **Localized perturbation** (including ligand displacement and ANM backbone motion), **Side Chain Sampling**, and **Minimization**. A bracket indicates that these three steps take 1-2 minutes to complete.

First thing: Ligand and protein check!

- Check Protein, missing loops, side chain...
- Ligand need to have unique atom names!
- Check protonation state, **read literature!**
- It needs to be one ligand (not composition of different ones...)
→ unique ligand number, name..
- Run the Protein Wizard of Maestro but check interactively each hydrogen bond, ligand, etc!!!

* LIGAND DATABASE FILE (OPLS2005)

*

```
INH  57  60  117  205  362
  1  0 M  CM  _C1_  1  46.079180 104.111990  50.326750
  2  1 M  CM  _C3_  3  1.338190  85.268590 104.753090
  3  1 S  CO4 _C5_  5  1.468930  37.845760 -103.947950
```

.....

```
11  16  8  4 10 18  3 18  2 10  1 13  4  3 19  4  3
12  3  4  5  1  2  1  1  2  1  1  2  1  1  8  6  4
```

.....

NBON

```
  1  3.5500  0.0760 -0.115000  2.0020  1.7750  0.023028004 -0.852763146
  2  3.5500  0.0760 -0.115000  2.0020  1.7750  0.023028004 -0.852763146
```

.....

BOND

```
  1  2  549.000  1.340
  1  3  320.000  1.462
```

.....

THET

```
  1  2  8  70.00000 124.00000
  1  2  9  35.00000 120.00000
```

....

PHI

```
  1  2  8  6 -0.28300  1.0  1.0
  1  2  8  6  0.26700 -1.0  2.0
```

....

IPHI

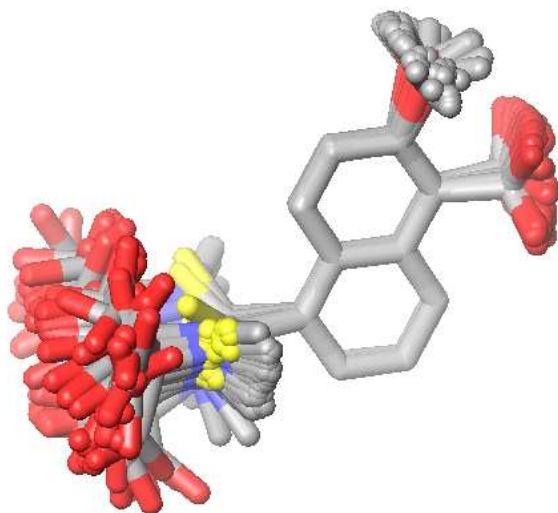
```
  3  4  1  2  1.10000 -1.0  2.0
  3  7  5  6  1.10000 -1.0  2.0
```

....

END

The **lig** file

The **LIG.rot.assign** file



```
rot assign res LIG &  
  sidelib FREE30 _C16 _C6_ &  
  sidelib FREE30 _C6_ _C22_ &  
  sidelib FREE30 _C22 _O5_ &  
  newgrp &  
  sidelib FREE10 _C10 _C11_ &  
  newgrp &  
  sidelib FREE10 _C16 _O4_ &
```

```
rot assign res LIG &  
  sidelib FREE30 _C16 _C6_ &  
  sidelib FREE30 _C6_ _C22_ &  
  sidelib FREE30 _C22 _O5_ &  
  sidelib FREE30 _C10 _C11_ &  
  sidelib FREE30 _C16 _O4_ &  
  sidelib FREE30 _C16 _O4_ &
```

Might the ligand dynamics be coupled to protein dynamics???

Global normal modes (INSPECT MODES AND MAYBE SELECT?):

.....

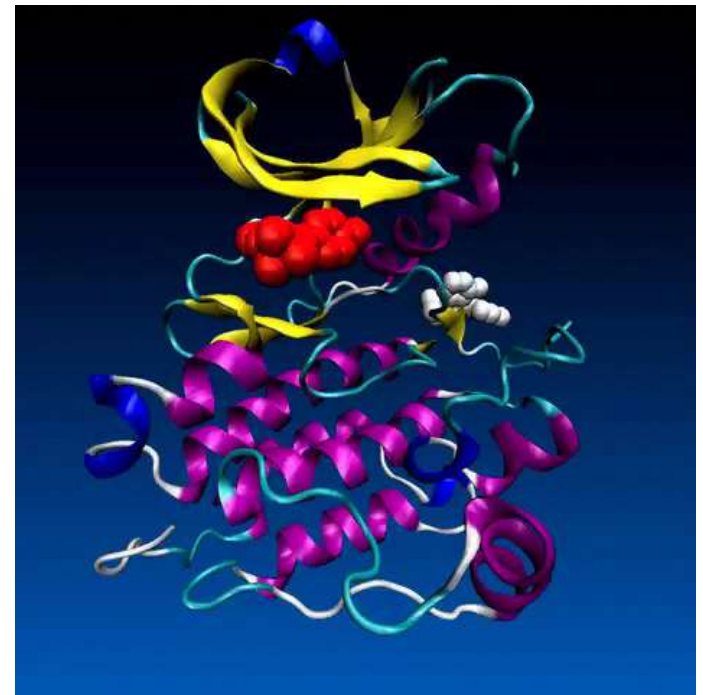
***caconst 0.1 &
anm_eig_freq 100000 &***

Specific mode! to open close some domain...

***anm_altm_freq 8 &
anm_altm_type 4 &
lanmanm mode 5 &***

general random case:

***anm_altm_freq 4 &
anm_altm_type 3 &***



And... avoid some highly moving parts to dominate the ANM motion!

```
caconst 0.01 &  
rem_bulk_mov 3 &  
anm_eig_freq 10000 &  
anm_altm_freq 6 &  
anm_altm_type 3 &  
lanmanm move_ca 0.75 &  
lanmanm neig 6 &  
lanmanm mix_modes 0.8 &  
lanmanm omit_no A:907 A:917 &  
lanmanm omit_no A:980 A:983 &
```


The control file

```
file datadir /home/bsc72/bsc72328/plop/data  
file log dyn.log
```

```
energy params solvent vdgbnp  
energy params ionic 0.15
```

```
load pdb INIT0001.pdb ions yes waters no het yes
```

```
constraint atom A:500:MG__ atom A:217:_OD1 200 1.9 &  
atom A:500:MG__ atom A:217:_OD2 200 1.9 &  
atom A:500:MG__ atom A:12:_OD1 200 1.82 &  
atom A:500:MG__ atom A:209:_OD1 200 1.82 &
```


```
load native 3vhv.pdb het yes
```

```
pele &  
het L:1 &
```

Focus more on concepts than on syntax!

```
file datadir /home/bsc72/bsc72328/plop/data  
file log dyn.log
```


```
energy params solvent vdgbnp  
energy params ionic 0.15
```



```
load pdb INIT0001.pdb ions yes waters no het yes
```



```
constraint atom A:500:MG__ atom A:217:_OD1 200 1.9 &  
atom A:500:MG__ atom A:217:_OD2 200 1.9 &  
atom A:500:MG__ atom A:12:_OD1 200 1.82 &  
atom A:500:MG__ atom A:209:_OD1 200 1.82 &
```



This will eliminate
the “abstraction of
the MG ion by the
ligand as we saw
before

```
load native 3vhv.pdb het yes
```



For comparison or
guiding!

```
pele &  
het L:1 &
```

```
pele &
  het L:1 &
  task &
    show bind_ene 1 &
    show rmsd_lig 1 heavy 0 &
    show atom 1 A:234:_CA_ &
    spawn point 1 -13.5 -3.6 76.8 lt 5.0 &
    if random 1 gt 0.5 then rot_r 0.1 else rot_r 0.25 endif &
  exit steps gt 2000 &
    exit point 1 -13.5 -3.6 76.8 lt 8.0 &
    exit atom 1 A:256:_CA_
  task &
    spawn point 1 -13.5 -3.6 76.8 within 8.0 &
    exit steps gt 500 &
end_task &
temp 1000 &
anmfreq 1 &
spfreq 1 &
mifreq 1 &
```

.....

wrfreq 1 &
path traj_ &
rem_bulk_mov 3 &
spradius 6 &
side &

Side options

randomize yes &
iter 1 &
sideend &
mirad 100 &
min &

Min options

rmsg 0.03 &
nbuf yes &
alphaup yes &
minimend &
caconst 0.1 &

ANM options

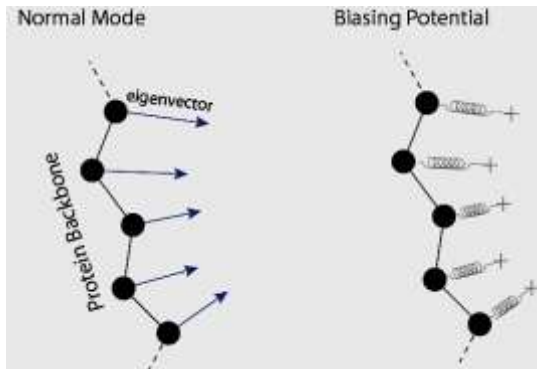
.....

anm_eig_freq 10 &
anm_altm_freq 6 &
anm_altm_type 3 &
lanmanm move_ca 0.75 &
lanmanm neig 6 &
lanmanm mix_modes 0.75 &

.....

lanmmin &
mxitn 100 &
iter 1 &
rmsg 0.02 &
nbuf yes &
alphaup no &
minimend

ANM-Min options



Free search

task &

```
....  
if random 1 gt 0.5 then rot_r 0.1 else rot_r 0.25 endif &  
if random 2 gt 0.5 then tra_r 6.0 tries 100 else tra_r 1.0 tries 5 endif &
```

end_task &

steered 1 & ??????

waitfor 3 & ??????

lcom_con 0.1 &

